International Journal of Zoology and Applied Biosciences Volume 2, Issue 6, pp. 292-298, 2017

https://doi.org/10.5281/zenodo.1312677

Research Article





A COMPARATIVE STUDY ON THE BIOCHEMICAL COMPOSITION OF HAEMOLYMPH OF THE FIFTH INSTAR LARVAE OF A MULTIVOLTINE BREED AND A BIVOLTINE BREED OF THE MULBERRY SILKWORM, BOMBYX MORI L.

*1Ponmurugan, M. and ²Karthikeyan, A.

¹PG and Research Department of Zoology, Government Arts College(Autonomous), Karur, Tamil Nadu, India

²PG and Research Department of Zoology, Government Arts College (Autonomous), Karur, Tamil Nadu, India

Article History: Received 8th November 2017; Accepted 19th November 2017; Published 21st November 2017

ABSTRACT

Changes in the composition of haemolymph reflect the physiological and biochemical transformations taking place in the insect tissues. In this work, the biochemical composition of haemolymph of the fifth instarlarvae of a multivoltine breed (L×CSR2) was compared with that of a bivoltine breed (CSR6×CSR12) of the mulberry silkworm, *Bombyx mori*. The larval period of the fifth instar larvae of both breeds lasted for 6 days. Increased trend of biochemical properties from the 1st day to the 6thday was noticed in both the breeds. The protein concentration of the fifth instar larvae of bivoltine breed was significantly lower than that of the multivoltine breed on all days. But the concentration of amino acid and carbohydrate were greater in the haemolymph of the bivoltine breed than that of the haemolymph of the multivoltine breed on all days. Lipid concentration was lower in the multivoltine breed than that of the bivoltine breed on the first four days whereas on the later days reverse trend was observed. The 't' values indicate that the differences in the biochemical parameters between the two breeds were significant on all days.

Keywords: Protein, Amino acids, Carbohydrate, Lipid.

INTRODUCTION

Insect haemolymph is a complex mixture of proteins. lipids, carbohydrates, amino acid, nucleic acids, hormones and their degradation products. It is primarily responsible for supplying nutrients, transferring metabolic wastes to maintain normal growth and development. It serves important roles in the immune system and in transport of hormones, nutrients, and metabolites. The silkworm has an open circulatory system containing haemolymph, which delivers nutrients and oxygen to all parts of the body. It is also an important repository for nutrition and energy. Major biomolecules such as proteins, carbohydrates and lipids play an important role in biochemical process underlying the growth and development of the silkworm (Ito and Horie, 1959). Haemolymph is the only extracellular fluid containing the products required for every physiological activity of the insect body. Thus changes composition of haemolymph reflect the physiological and

biochemical transformations taking place in the insect tissues. Lauffer (1960) was the first ever observed haemolymph proteins in silkworm Bombyx mori. Haemolymph protein content increases throughout the fifth instar and reaches maximum at the end of the fifth instar in the silkworm races. Murthy et al. (2014) reported that the total proteins in the whole body rapidly increases from the 1st instar and reaches maximum at the end of the 4th instar in multivoltine (Pure Mysore PM), crossbreed (PM×CSR2) and bivoltine (CSR2) breeds of the silkworm. Haemolymph proteins and carbohydrates rapidly increase from the 1st instar and reach maximum at the end of the 5th instar. Only a very little information pertaining to variations in the composition of the haemolymph of different races of silkworm is available. Thus a comparative analysis on changes in the major biochemical constituents such as carbohydrates, proteins, amino acids and lipids in the haemolymph of the fifth instar larvae of two breeds of the mulberry silkworm, *Bombyx mori* i.e. L×CSR2 and CSR6×CSR12 was carried out in this work.

MATERIALS AND METHODS

Silkworm breeds used for study

In the present study two races of the commercially exploited multivoltine crossbreedL×CSR2 and Bivoltine breed CSR6×CSR12 silkworm *B. mori* were selected. The eggs of this race were procured from Silkworm Rearing Department, Rayanur, Karur district. They were kept under lab conditions and allowed to hatch. The emerged first instar larvae were fed with young leaves of mulberry variety MR2. The larvae were acclimatized to the lab conditions by rearing them during the month of November, 2016 till fifth instar in the laboratory with the relative humidity of 80-90% and the temperature of 27-30°C.

Haemolymph collection

Haemolymph was collected in a pre-chilled eppendorf tube containing a few crystals of thiourea by cutting the first proleg of fifth instar larva. Haemolymph was collected at an interval of 24h for six days from day one. Haemolymph was centrifuged at 3000 rpm for 3 min and supernatant was used for estimation of total haemolymph ptoteins, amino acids, carbohydrates and lipids.

Estimation of Protein, Carbohydrate, Amino acid and Lipid

Total haemolymph protein was estimated according to Lowry *et al.* (1951) using Bovine Serum Albumin (BSA) as standard. The quantitative estimation of total carbohydrate in haemolymph was done by anthrone method of Dubois *et al.* (1956) using glucose as standard. Total haemolymph amino acid was estimated by Ninhydrin method of Moore and Stein, (1954) using aspartic acid and leucine as standard. Total haemolymph lipids was estimated by the method of Zoellner and Kirsch (1962) using cholesterol as standard. The results were statistically analyzed and discussed.

RESULTS AND DISCUSSION

Haemolymph brings about functional homeostasis of insect organs by transporting chemical substances into and out of the cells of the tissues and thus serves as a medium of chemical communication of distant and distinct organ systems of insect body. It serves as the transport milieu for the exchange of essential materials between cells, tissues and organs (Mullins, 1985; Karpells *et al.*, 1990). Transport of biochemicals from the different tissues may be required to meet the higher physiological activities in silkworm such as increased body growth and cocoon formation. So the biochemical parameters such as, proteins, amino acids, carbohydrates, lipids, nucleic acids etc., vary significantly during the life cycle of all living organisms. The quantitative variation of these biomolecules in insects during growth and metamorphosis have been reported by

Nagata and Yashitake (1989). Similarly quantitative variations in the concentration of protein, amino acids and carbohydrates of the haemolymph have been observed on all days of the fifth instar larvae of both breeds of the mulberry silkworm in the present study (Tables 1, 2, 3 & 4). The results of the present work were in line with the report of Yogananda Murthy (2015) that is the bivoltine breed was superior to multivoltine with respect to amino acid and carbohydrate contents of the haemolymph. But higher content of protein was noticed on all days in the haemolyph of the multivoltine breed compared to that of the bivoltine breed. Quantity of lipid was greater in the haemolyph of the bivoltine than that of the multivoltine breed till 4th day and this condition reversed on the 5th and 6th days. 't' values of all parameters on all days were significant at p < 0.05. This indicated that the differences between the biochemical compositions of haemolymph of two breeds were greater and significant. Such significant differences between the two breeds might be correlated with the different in the inherent utilization ability. The utilization capacity of bivoltine must be greater as the haemolymph protein concentration in bivoltine was always lower than that of the multivoltine larvae.

Proteins

In this study, the larval period of the fifth instar of the larvae of both breeds lasted for 6 days. In multivoltine, protein content of haemolymph was found to be increasing during the larval period of the fifth instar from day one to the sixth day. On the first day, haemolymph contained 167.5 mg/ml of protein. It increased to 205.6 mg/ml on the sixth day. In bivoltine breed also, an increase in protein was observed from 63.6 mg/ml on the first day to 149.8 mg/ml on the last day. Comparatively, the protein concentration of the haemolymph of the fifth instar larvae of bivoltine breed was significantly lower than that of the multivoltine breed on all days.

Proteins play an important role in the growth and development of B. mori and synthesis of silk proteins in silk gland during larval development (Seo et al., 1985). During the larval development high molecular weight proteins are synthesized in large amount by the larval fat body and secreted into the haemolymph. Such proteins are known as major larval haemolymph proteins or storage proteins. Synthesis of such proteins is dependent on the silkworm during the larval nutritional status of development (Ramesh Babu etal., 2009) environmental conditions (Benchamin and Anantharaman, 1990; Ramesha et al., 2010). Proteins are important for the development, metamorphosis and to maintain a number of physiological functions (Murthy et al., 2014). Variations in haemolymph proteins reflect the balance between the synthesis, storage, transport and degradation of structural and functional proteins during ontogeny as well as response to particular ecological and physiological conditions (Florkin and Jeuniaux, 1974).

Haemolymph proteins undergo radical changes both in quality and quantity during development. In the present study the concentration of protein in haemolymph increased progressively during the larval development and reached maximum in the late fifth instar larvae of both races as recorded by Lauffer (1960) in three races, multivoltine (Pure Mysore PM), crossbreed (PM×CSR2) and bivoltine (CSR2). It was found that during the development of the fifth instar larvae of Samiaricini, there is a rapid increase in the haemolymph protein concentration which attains its peak at the end of larval life irrespective of the season. Similar observations have been recorded where protein concentration was found to be higher at the end of larval stage in the silkworm (Ito and Arai, 1963; Banno et al., 1993; Murthy et al., 2014). The increase in feeding and growth as reported by Hurlimann and Chen, (1974) and active secretion of proteins and carbohydrate by other tissues like fat bodies as proved by Nagata and Kobayashi. (1990) and Chen (1978) could be cited as reasons. The increasing trend was to meet the higher requirement of these macromolecules during metamorphosis. concentration of haemolymph proteins could be correlated with high consumption of mulberry leaves subsequently higher rate of conversion and their accumulation in haemolymph (Banno et al., 1993; Aruga, 1994).

In the present study, a steady increase in the haemolymph protein concentration during the development of last larval instar indicates the higher utilization of dietary proteins. The increase in protein could be attributed to the regular feeding, initiation of silk protein synthesis in silk gland at end of the fifth instar and the development of reproductive organs, and the increased rate of metabolism (Sinha and Sinha, 1994). Such increased proteins could be the compensatory replacement of the proteins which are utilized for the formation of puparium (Malik and Malik, 2009). The most abundant proteins in larval haemolymph belongs to a class known as storage or hexamerins which are synthesized by the fat body and reach extremely high concentrations in the last instar (Sumino et al., 1980; Kanost et al., 1990). These results were in confirmation with the earlier works of Satish (1998) that, haemolymph protein in sericigenous insects is responsible for the formation of silk proteins in silk glands. High protein concentration in fifth instar larval was an indication of a greater metabolic activity and immune response. The haemolymph of the multivoltine breed was found to have higher protein level than that of the bivoltine breed and thus it was considered as a better strain in terms of cocoon weight and shell weight as reported by Chakravorty and Neog (2006). On contrary Banno et al. (1993) reported that the bivoltine showed a higher protein content in the haemolymph followed by cross breed and multivoltine during fifth instar. High protein concentration is an indication of greater metabolic activity. Synthesis and utilization of haemolymph proteins are conditioned by genetic and hormonal control (Hurliman and Chen, 1974).

Amino acids

The quantity of amino acid ranged from 44.0 mg/ml on the first day to 140.0 mg/ml on the sixth day in the multivoltine breed. In Bivoltine breed (CSR6×CSR12),

fifth instar larvae recorded an increase in the amino acid level from 122.9 mg/ml on the first day to 176.6 mg/ml on the first day to 149.8mg/ ml on the last day. Though the increasing trend was noticed in the amino acid content of the haemolymph of both breeds, the increase was only gradual in the bivoltine breed whereas it was highly significant in the multivoltine breed. Amino acids are the essential components of all living cells. They are the building blocks of proteins. Many insects including the silkworm are known to contain usually large amount of free amino acids (Ramsay, 1958). Amino acids are essential for growth and survival of the silkworm. Shimura (1978) reported that, haemolymph acts an amino acid reservoir between midgut and silk gland and supplies amino acids to silk gland for silk synthesis. The amino acid content of the haemolymph was found to be gradually increasing from the first day to last day. Highest level of amino acid was recorded on the fifth day and the increase in amino acid could be attributed to the initiation of protein balance in the silkworm at end of the fifth instar and the proteolysis. The amino acid level of bivoltine race is higher than the multivoltine race silkworm B. mori. In the present investigation variations in the amino acid levels of two breeds were the reflections of the variations in the protein metabolism of two breeds which are ultimately the result of genetic variations. The total free amino acid amino acid levels declined from the early-fourth instar to the mid-fifth instar and were elevated during the late-fifth instar in the metamorphosing silkworm, B. mori (Sivaprasad and Muralimohan, 1990). The differences in the amino acid composition between two breeds reflected in the quality of silk they produced. Such difference was ultimately due to genetic variation.

Carbohydrates

The quantity of carbohydrate ranged from 8.2 mg/ml on the first day to 35.0 mg/ml on the sixth day in the multivoltine breed. In Bivoltine breed (CSR6×CSR12), fifth instar larvae recorded an increase in the carbohydrate level from 34.1mg/ml on the first day to 117.0 mg/ ml on the last day. The percentage of increase in carbohydrate was not uniform in multivoltine breed whereas the increase in bivoltine breed was high and gradual till the fourth day and then the increase was very low on later days (6.35 and 12.7%). Carbohydrates also play an important role as energy source and protecting silkworm during adverse condition. The late age silkworm larvae accumulate higher carbohydrates compared to young age worms. Simex and Kodrik (1986) have reported that the free carbohydrate in the haemolymph changed significantly during last larval instar of the silkworms. The level of carbohydrates during larval development reveals the degree of utilization of carbohydrates, which are the major sources of energy in the body, for growth and development of the larva that might ultimately determine the difference in the quality and quantity of silk production. The carbohydrate content of the haemolymph was found to be gradually increasing from the first day to last day. The total sugar content in the haemolymph increased constantly from the first to sixth day in bivoltine race. Glycogen and trehalose are main constituents of haemolymph and they play an important role during growth, metamorphosis and diapause (Jo and Kim, 2001). It has been reported that high concentration of carbohydrates in hemolymph are maintained during larval development as energy reserve to be utilized later during metamorphosis, pupal and adult stage (Simex and Kodrik, 1986; Mishra et al., 2010). Higher carbohydrate in the bivoltine breed revealed that it had the higher capacity accumulate carbohydrates than the multivoltine breed. Carbohydrates increased with the advancement of age of larva and reached at its peak on the last day of fifth instar larva of both breeds. Similar observation has been recorded by Simex and Kodrik, (1986), Mishra et al. (2010) and Murthy et al. (2014). Higher concentration of carbohydrates in haemolymph of later stages of development could be utilized for the energy required for the formation of cocoon, pupal and adult cuticle and other developmental processes (Simex and Kodrik, 1986; Misra et al., 2010; Chippandale, 1978).

Lipids

The quantity of lipids ranged from 6.66 mg/ml on the first day of the fifth instar to 44.44 mg/ml on the sixth day in the multivoltine breed. In Bivoltine breed (CSR6×CSR12), fifth instar larvae recorded an increase in the lipid level from 30.24 mg/ml on the first day to 35.57 mg/ml on the last day. Gradual increase was noticed in the carbohydrate content of bivoltine breed whereas about 133% and 60% increase was noticed on the second and fifth day respectively of the multivoltine breed. The total lipid

respectively of the multivoltine breed. The lipidcontent was found to be increasing as the day advanced in the haemolymph of fifth instar larvae of both breeds of the silkworm. However lower level of lipids was observed in multivoltine variety than the bivoltine variety on the first four days and later reverse trend was observed on the 5^{th} and 6^{th} days. There was a significant difference between the two breeds in the lipid level of haemolymph on all days. Lipids are used as a source of energy required for growth and metamorphosis of insects. The lower level of lipid in the multivoltine breed might be due to the over utilization of lipid and fatty acids for growth, moulting and metamorphosis (Mallikarjuna et al., 2016). Mobilization of lipid to the fat body might also be and increased lipase activity the reasons for low level of lipids in multivoltine breed (Streit, 1978).

The 't' values indicate that the differences in the biochemical parameters between the two breeds were significant on all days. The differences between the multivoltine and bivoltine breeds of silkworm in terms of major biochemical composition in the haemolymph might be attributed to the differences in feeding efficiency, utilization efficiency, conversion efficiency, hormones level, metabolic rates etc. Though many physiological reasons were cited, the differences are breed specific and their different adaptability are attributed to the genetic characteristics gained from their parental stock. Thus biochemical profile of haemolymph can be used as an index to screen germplasm stock for developing a breed with higher survival rate.

Table 1. Quantity of Protein in the haemolymph of the fifth instar larvae of a multivoltine breed (L×CSR2) and a bivoltine breed (CSR6×CSR12) of *Bombyx mori*.

	Multivoltine	Bivoltine	't'- test value
Days	L×CSR2 (mg/ml)	CSR6×CSR12 (mg/ml)	Level of Significance at p < 0.05
1 st day	167.57±1.67	63.71± 1.27	110.18 Significant
2 nd day	$171.02 \pm 1.57 (2.0\%)$	66.29 ±1.29 (4.0%)	114.87 Significant
3 rd day	$180.59 \pm 1.14 (5.5\%)$	$82.75 \pm 1.10 \ (24.9\%)$	137.30 Significant
4 th day	$189.14 \pm 1.01 (4.7\%)$	$105.19 \pm 1.0 \ (27.0\%)$	131.45 Significant
5 th day	199.11 ±1.21 (5.2%)	121. 60 ±0.89 (15.6%)	115.44 Significant
6 th day	205.64 ±1.29 (3.2%)	$149.84 \pm 1.82 \ (23.1\%)$	55.86 Significant

Values inside the parentheses indicate the percentage of increase in biochemical content over the previous day.

Table 2. Quantity of Amino acids in the haemolymph of the fifth instar larvae of a multivoltine breed (L×CSR2) and a bivoltine breed (CSR6×CSR12) of *Bombyx mori*.

	Multivoltine L×CSR2	Bivoltine CSR6×CSR12	't'- test value
Days	(mg/ml)	(mg/ml)	Level of Significance at p < 0.05
1 st day	44 ±1.22	122.90 ± 1.54	-89.33 Significant
2 nd day	$80 \pm 1.58 (81.8\%)$	$127.27 \pm 1.63 (3.4\%)$	-46.53 Significant
3 rd day	94 ±1.58 (17.5%)	$135.63 \pm 1.74 (6.6\%)$	-38.75 Significant
4 th day	$110 \pm 1.58 \ (17.0\%)$	$157.09 \pm 0.88 (11.2\%)$	-58.12 Significant
5 th day	122 ±1.58 (10.9%)	$172.90 \pm 3.21 (14.5\%)$	-31.68 Significant
6 th day	$140 \pm 1.58 (14.7\%)$	$176.67 \pm 1.74 (2.1\%)$	-34.82 Significant

Values inside the parentheses indicate the percentage of increase in biochemical content over the previous day.

Table 3. Quantity of Carbohydrate in the haemolymph of the fifth instar larvae of a multivoltine breed (L×CSR2) and a bivoltine breed (CSR6×CSR12) of *Bomby xmori*.

	Multivoltine	Bivoltine	't'- test value
Days	L×CSR2	CSR6×CSR12	Level of Significance at $p < 0.05$
	(mg / ml)	(mg / ml)	
1 st day	8.21±0.54	34.17 ± 0.64	-66.04 Significant
2 nd day	11.78±0.73 (42.6%)	47.01±1.58 (37.8%)	-44.99 Significant
3 rd day	13.57±1.47 (15.3%)	65.22± 2.28 (38.72%)	-42.39 Significant
4 th day	20.71±0.72 (53.3%)	97.61± 1.56 (49.6%)	-99.81 Significant
5 th day	22.50±1.17 (8.6%)	103.88±1.60 (6.35%)	-91.31 Significant
6 th day	35±1.58 (55.5%)	117.01±1.22 (12.7%)	-91.67 Significant

Values inside the parentheses indicate the percentage of increase in biochemical content over the previous day.

Table 4. Quantity of Lipids in the haemolymph of the fifth instar larvae of a multivoltine breed (L×CSR2) and a bivoltine breed (CSR6×CSR12) of *Bombyxmori*.

	Multivoltine	Bivoltine	't'- test value
1 st day	6.66 ± 0.47	30.24 ± 0.90	-51.45 Significant
2 nd day	$15.55 \pm 0.15 \; (133.4\%)$	$31.57 \pm 0.72 \ (4.3\%)$	-48.04 Significant
3 rd day	$17.77 \pm 0.10 \ (14.2\%)$	$32.09 \pm 0.52 \; (1.6\%)$	-59.79 Significant
4 th day	$22.22 \pm 1.10 \ (25.0\%)$	$32.24 \pm 0.81 \; (0.46\%)$	-16.34 Significant
5 th day	$35.55 \pm 0.67 \ (59.9\%)$	$33.19 \pm 1.07 \ (2.9\%)$	4.15 Significant
6 th day	$44.44 \pm 0.67 \ (25.0\%)$	$35.57 \pm 0.85 \ (7.17\%)$	15.26 Significant

Values inside the parentheses indicate the percentage of increase in biochemical content over the previous day.

CONCLUSIONS

Quantitative variations in the concentration of protein, amino acids and carbohydrates in the haemolymph have been observed on all days of the fifth instar larvae of both breeds of the mulberry silkworm. The concentration of protein in haemolymph increased progressively during the larval development and reached maximum in the late fifth instar larvae of both races. The multivoltine breed was found to have higher protein level in the haemolymph of the fifth instar. The concentration amino acid and carbohydrate were greater in the haemolymph of the bivoltine breed than that of the haemolymph of the multivoltine breed on all days. Lipid concentration was lower in the multivoltine breed than that of the bivoltine breed on the first four days whereas on the later days reverse trend was observed. The differences between the multivoltine and bivoltine breeds of silkworm in terms of major biochemical composition in the haemolymph are breed specific and are attributed to their genetic characters.

ACKNOWLEDGEMENTS

We wish to express sincere thanks to the Principal and Head and other staff of the PG and Research Department of Zoology of Government Arts College (Autonomous), Karur, Tamil Nadu for providing the laboratory facilities to carry out this research work.

REFERENCES

Aruga, H., 1994. Principles of Sericulture. New Delhi: Oxford IBH Publishing Co. Pvt. Ltd, New Delhi, India, pp.376.

Banno, Y., Tochihara, S., Kawaguchi, Y. and Doira, H., 1993. Protein of young larva of silkworm *Bombyx mori* L. *J. Sric. Sci. Japan.*, 62(3), 187-194.

Benchamin, K.V. and Jolly, M.S., 1986. Principles of silkworm rearing, Proceedings of seminar on Prospects and problems of Sericulture. Edi. S. Mahalingam, Madras, pp 63-108.

Benchamin, K.V. and Anantharaman, K.V., 1990. Standardisation of moulting test to evaluate mulberry leaf quality under tropical conditions. *Ind. J. Seric.*, 29 (2), 255-256.

Chakravorty, R. and Neog, K., 2006. Food plants of eri silkworm *Samiaricini* (DONOVAN) their rearing performance and prospects for exploitation. Leeds paper of National workshop on eri food plants, Held at Guwahati, 11th-12th October.

- Chen, P.S., 1978. Protein synthesis in relation to cellular activation and deactivation. In: Biochemistry of Insects (ed. Rockstein, N.). Academic Press, Newyork, pp.145-200.
- Chippandle, G.M., 1978. The function of carbohydrates in insect life precesses. In: Biochemistry of insects (ed. Rockstein, N.) Academic Press, Newyork, pp. 2-54.
- Dubois, M., Gilles, K.A., Hamilton, J.K., Rebers, P.A. and Smith, F., 1956. Colorimetric method for determination of sugars and related substances. *Anal. Chem.*, 28(3), 350-356.
- Florikin, M. and Jeuniaux, C., 1974. Hemolymph composition. In: The Physiology of Insects (Edited by Rockstein, M.) Vol. 5, pp. 255-307, Academic Press, New York.
- Hurliman, R.F. and Chen, P.S., 1974. Ontogenetische Veranderungen des enzymmusters in der haemolymphe von Phormiarigina. *Revue. Swisse. Zool.*, 81, 648-654.
- Ito, T. and Arai, N., 1963. Food values of mulberry leaves for the silkworm *Bombyx mori* L. determined by means of artificial diets. I. Relationship between kinds of mulberry leaves and larval growth. *Bull. Seric. Exp. Stn. Japan.*, 18(4), 226-229.
- Ito, T. and Horie, Y., 1959. Carbohydrate metabolism of the midgut of the silkworm, *Bombyx mori* L. *Arch. Biochem. Biophys.*, 80, 174-176.
- Jo, M.H. and Kim, Y., 2001. Relationship between cold hardiness and diapause in the small fruit Tortix, Adoxophyes orana (Fischer Von Roslerstamm). J. Asia Pacific. Entomol., 4(1), 1-9.
- Kanost, M.R., Kawooya, J.K., Law, J.H., Ryan, R.O., Van Heusden, M.C. and Ziegler, R., 1990. Insect haemolymph proteins. *Advan. Insect Physiol.*, 22, 299-396.
- Karpells, S.T., Leonard, D.E. and Kunkel, J.G., 1990. Cyclic fluctuations in arylphorin, the principal serum storage protein of *Lymantria dispar*, indicate multiple roles in development. *Insect Biochem.*, 20, 73-82.
- Krishnaswamy, S., 1978. New technology of silkworm rearing. Bulletin No. 2, pp. 1-24. Central Silk Board, Mysore, India.
- Lauffer, H., 1960. Blood proteins in insect development. Ann. New York Acad. Sci.., 89, 415-490.
- Lowry, O.H., Rosebrough, N.J., Farr, A.L., Randall, R.J., 1951. Protein measurement with folin-phenol reagent. *J. Biol. Chem.*, 193, 265-275.

- Malik, M.A. and Malik, F.A., 2009. Ontogenic Changes in Haemolymph biochemical composition in the silkworm, *Bombyx mori* L. under thermal stress. *Acad. J. Entomol.*, 2 (1), 16-21.
- Mallikarjun, G., Neetha, N.K., Manjunatha, B., Sivaprasad, V. and Shyam Kumar, V., 2016. A mini review of functional proteins in silkworm *Bombyx mori L.* haemolymph. *Indian J. Sci. Technol.*, 9(38), 1-8.
- Mishra, P.K., Kumar, D., Jaiswal, L., Kumar, A., Singh,
 B.M.K., Sharan, S.K., Pandey, J.P. and Prasad, B.C.,
 2010. Biochemical aspects of diapause preparation in ultimate instar of tropical tasar silkworm *Antheraea* mylitta Drury. J. Ecophysiol. Occupat. Health, 9, 253-260
- Moore, S. and Stein, W.H., 1954. A modified ninhydrin reagent for the photometric determination of Amino acids and related compounds. *J. Biol. Chem.*, 211(2), 907-913.
- Mullins, D.E., 1985. Chemistry and physiology of the hemolymph. In: Comprehensive Insect Physiology Biochemistry and Pharmacology (Edited by Kerkut, G. A. and Gilbert, L.I.), Vol. 3, pp. 355-400, Pergamon Press, Oxford.
- Murthy, V.N.Y., Ramkumar, B., Jayaram, G.N. and Lokesh, G., 2014. Critical biochemical analysis in different body tissues in three commercial silkworm (*Bombyx mori* L.) races. *As. J.N. App. Sci.*, 3(2), 20-30.
- Nagata, M. and Kobayashi, M. 1990. Quantitative changes in the storage proteins during larval development of silkworm *Bombyx mori* L. *J. Seric. Sci. Japan.*, 59(6), 461-468.
- Ramesh Babu, K., Ramakrishna, S., Reddy, Y.H.K., Lakshmi, G., Naidu, N.V., Basha Sadak, S. and Bhaskar, M. 2009. Metabolic alteration and molecular mechanism in silkworm larvae during viral infection, A review. *Afri. J. Biotech.*, 8(6), 899-907.
- Ramesha, C., Anuradha, C.M., Lakshmi, H., Sugnana Kumari, S., Seshagir, S.V., Goel, A.K. and Suresh Kumar, C., 2010. Nutrigenetic traits analysis for identification of nutritionally efficient silkworm germplasm breeds. *Biotechnology*, 9, 131-140.
- Satish, B.R., 1998. Improvement of nutritive status of mulberry to increase silk production. Ph.D. Thesis, Bangalore University, Bangalore, India, pp.108-146.
- Seo, R.W., Youn, C.Y., Kang, C.S. and Kim, H.R., 1985. A study on protein pattern of haemolymph during last larval and pupal stages of *Bombyx mori* L. *Bull. Entomol. Res.*, 11, 153-164.

- Shimura, K. 1978. Synthesis of silk proteins. *Silkworm:* An important laboratory tool (Ed. T. Tazima). Kodansha Scientific Books, Kodensha Ltd. Japan, pp. 189-211.
- Simex, V. and Kodrik, D., 1986. Changes in the tissue glycogen and free carbohydrates of haemolymph during the last larval instar and metamorphosis of silkworm, *Bombyx mori L. Acta, Entomol. Bohemaslov.*, 83(2), pp 92-100.
- Sinha, AK., Sinha, USP., Sinha, SS. and Sengupta, K., 1994. Studies on free amino acids, proteins carbohydrates and phosphorus compounds in the tissue extracs of healthy and pebrine infected moths of Tasar silkworm *Antheraea mylitta D. Ind. J. Seric.*, 30, 103-106.
- Sivaprasad, S. and Muralimohan, P., 1990. Amino acids,

- aminotransferases and proteins in the metamorphosing silkworm, *Bombyx mori* L. *Proc. Ind. Acad. Sci. Anim. Sci.*, 99, 369-375.
- Streit, B., 1978. Changes in protein lipid and carbohydrate during starvation in the freshwater limpet *Ancylus fluviatilis* (Basommatophora). *J. Comp. Physiol.*, 123, 149-153.
- Sumino, T., Masao, N. and Masahiko, K., 1980. Storage proteins in the silkworm *Bombyx mori* L. *Insect Biochemistry.*, 10, 289-303.
- Yogananda Murthy, V.N., 2015. Estimation of protein concentration in different tissues of popular silkworm *Bombyx mori* L. *Int. J. Adv. Res.*, 3(1), 254-261.
- Zoellner, N., and Kirsch, K., 1962. Colorimetric method for determination of total Lipids. *J. Exp. Med.*, 135, 545-550.