

## INNATE IMMUNE SYSTEM AND MOLECULAR ELEMENTS IN FRESHWATER CRAB: A MINI REVIEW

<sup>1,2\*</sup>Mahadev Asaram Jadhav and <sup>2</sup>Encily Reymend Martin

<sup>1\*</sup>Department of Biotechnology & Bioinformatics, Deogiri College (Autonomous), Chhatrapati Sambhajnagar- 431 005

<sup>2</sup>Department of Zoology, Dr. Babasaheb Ambedkar Marathwada University, Chhatrapati Sambhajnagar-431 001

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### ABSTRACT

The immune system of freshwater crabs is an intricate network essential for their survival in pathogen-rich environments. This review synthesizes current knowledge on the molecular mechanisms underlying immune responses in the hemolymph of freshwater crabs. Key components such as hemocytes, antimicrobial peptides, and the prophenoloxidase system are discussed alongside recent advancements in molecular techniques that have facilitated these discoveries. We review some basic aspects of crab effector defense processes, like agglutination, encapsulation, phagocytosis, clottable proteins, and bactericidal activity, induced by these carbohydrate-driven recognition patterns. Understanding these immune defense mechanisms provides insights into invertebrate immunity and has practical implications for aquaculture.

**Keywords:** Hemolymph, Immune defense, Hemocytes, Antimicrobial peptides, Prophenoloxidase system.

### INTRODUCTION

Freshwater crabs inhabit environments with diverse pathogenic threats, relying primarily on their innate immune system for defense. Unlike vertebrates, crabs lack an adaptive immune system, making the study of their innate immune responses crucial for understanding their survival strategies. Immunity studies in vertebrate and invertebrate species suggest that the invertebrate defense mechanisms could be considered precursors of vertebrate immunity. These events require the participation of cellular groups and humoral factors generated against specific antigens, as has been shown in mammals (Hoffman *et al.*, 1999; Iwanaga and Lee, 2005). In invertebrates, there is little evidence identifying whether any of these factors are antigen-specific. However, some cellular and cell-free hemolymph factors show high specificity for non-self or damaged cells, similarly as demonstrated for antibodies (Yeaton, 1981; Vasa *et al.*, 1999). Crustaceans represent a group of economic relevance, because of their adaptation to aquaculture; however, crustaceans are affected by diseases caused, mainly, by opportunistic pathogens causing huge economic losses. Studies on the immunity of invertebrates have focused on identifying defense mechanisms and biochemical pathways activated during an infection, and on

identifying cell-free hemolymph and cellular factors involved in the destruction of pathogens, regulation, and damage repair. Crustaceans possess an open circulatory system, where nutrients, oxygen, hormones, and cells are distributed in the hemolymph. The hemocytes (circulating cells) could be functionally analogous to vertebrate leukocytes, because they are mainly involved in non-self-matter recognition and elimination (Sritunyaluksana and So'derha'll, 2000), as well as in downstream coagulation (Cerenius *et al.*, 1994; Vázquez *et al.*, 1997).

Immune response in hemolymph of crabs is triggered by pathogen-associated molecular patterns (PAMPs) present on surface of microbes, such as lipopolysaccharides (LPS) of Gram-negative bacteria,  $\beta$ -1,3-glucans of fungi, and peptidoglycans of Gram-positive bacteria (Chen *et al.*, 2002). Generally, PAMPs are recognized via a set of pattern-recognition receptors (PRRs) that are germline-encoded receptors of the innate immune system (Ariki *et al.*, 2004). In horseshoe crabs, there are some special serine proteases in granular hemocytes, including Prochelicerae C, Prochelicerae B, and Prochelicerae G, which can directly recognize LPS and  $\beta$ -1,3-glucans and activate the innate immune response (Kobayashi *et al.*, 2015; Muta *et al.*, 1995). However, the cell-surface receptors in horseshoe

\*Corresponding Author: Mahadev Asaram Jadhav, Department of Biotechnology, Deogiri College, Chhatrapati Sambhajnagar-431005. Email: majadhav22g@gmail.com

crabs have not been identified. In addition, a Toll-like protein named tToll has been identified in horseshoe crab hemocytes and shown to be expressed in multiple tissues (Inamori, 2004). Interestingly, there is no evidence to demonstrate that the tToll protein is involved in pathogens recognition. Obviously, the innate immune mechanism of horseshoe crabs is unique and complex, especially against Gram-negative bacteria containing LPS in the cell wall membrane. In previous studies, many innate immunity-related genes have been reported in Chinese horseshoe crab (Shibata *et al.*, 2018), but the immune response mechanism based on high-throughput analysis of pathogen challenge has been rarely reported.

Antimicrobial peptides or substances are the host defense compounds that have recently drawn attention, due to their properties and diversity. In crustaceans, these substances are considered to be a main component of innate immunity (Smith and Chisholm, 2001). There are few reports evaluating the bioactivity of crustaceans, and many researchers have studied the antibacterial activity of marine crustaceans and prawns (Tonganunt *et al.*, 2008; Stewart and Zwicker, 1972; Noga *et al.*, 1996; Khoo *et al.*, 1999; Ravichandran *et al.*, 2010). The crabs are the rich sources of bioactive compounds, but the researchers carried out the pharmacological properties of marine crabs (Veeruraj, *et al.*, 2008; Anbuhezien and Ravichandran, 2009) but not in freshwater crabs. Hence, the present study was aimed to investigate the antimicrobial potency of haemolymph collected from freshwater crab, *Paratelphusa hydrodromous*.

### Defense mechanisms in crustaceans

In invertebrates, the physical barriers are the first obstacle to detain pathogenic micro-organisms (Söderhäll, 1982). When there is damage and the micro-organisms invade the tissue, proteolytic pathways take place instantly, allowing elimination or diminution of microbes invading the organism (Ratcliffe *et al.*, 1985). The effector mechanisms for invertebrate immune responses include the coagulation cascade, which avoids the loss of hemolymph and stimulates oxidative metabolites and production of melanin by activating the proPO system (Sritunyalucksana and Sodarhall, 2000; Vargas-Albores and Yepiz-Plascencia, 2000; Kawabata *et al.*, 1996). Prophenoloxidase activation stimulates other important processes in the immune response, such as phagocytosis, encapsulation, and nodule formation (Söderhäll *et al.*, 1990; Smith and Chisholm, 2001). Activation of such processes seems to be mediated through the specific recognition of glycosylated pathogen-associated molecular patterns (PAMPs) by crustacean proteins. This review focuses on the molecular characterization of immune defense mechanisms in the hemolymph of freshwater crabs, summarizing the roles of various immune components and highlighting recent research advancements.

### Hemolymph composition and immune function

The hemolymph of freshwater crabs is analogous to the blood of vertebrates, containing immune cells (hemocytes)

and soluble factors that play critical roles in pathogen defense.

### Hemocyte types and functions

Hemocytes are categorized into three main types:

1. Granulocytes: Contain granules with antimicrobial substances and are involved in degranulation.
2. Hyalinocytes: Predominantly phagocytic cells that ingest and degrade pathogens.
3. Semigranulocytes: Participate in encapsulation and nodule formation, trapping larger pathogens (Johansson *et al.*, 2000; Söderhäll *et al.* 1983).

### Soluble immune factors

The hemolymph also contains various soluble factors, including antimicrobial peptides (AMPs) and components of the prophenoloxidase (proPO) system, which are crucial for humoral immune responses (Cerenius and Söderhäll, 2004; Amparyup *et al.*, 2008).

### Pathogen recognition mechanisms

Pathogen recognition in freshwater crabs is mediated by pattern recognition receptors (PRRs) that detect pathogen-associated molecular patterns (PAMPs).

#### Pattern Recognition Receptors

Key PRRs include:

- a. Toll-like Receptors (TLRs): Recognize a broad range of PAMPs and activate signaling pathways that induce immune responses.
- b. Lectins: Bind to specific carbohydrate structures on the surfaces of pathogens, facilitating their recognition and clearance (Iwanaga and Lee, 2005; Rozen and Skaletsky, 2000).

### Cellular immune responses

Hemocytes execute various cellular immune responses, including:

1. Phagocytosis: Hyalinocytes engulf and digest pathogens.
2. Degranulation: Granulocytes release antimicrobial compounds from their granules.
3. Encapsulation: Semigranulocytes form capsules around larger pathogens to isolate and neutralize them (Shields, 2003; Cerenius *et al.*, 2008).

### Humoral immune responses

Humoral responses involve the production of AMPs and activation of the proPO system.

### Antimicrobial peptides (AMPs)

AMPs, such as crustins and defensins, are produced in response to pathogen recognition and play a direct role in neutralizing pathogens by disrupting their cell membranes (Lee *et al.*, 2006).

### Prophenoloxidase system

The proPO system is activated upon pathogen recognition, leading to melanization, which encapsulates and immobilizes pathogens, preventing their spread (Johansson *et al.*, 2000; Cerenius and Söderhäll, 2004).

### Signal transduction pathways

Activation of immune responses involves several signal transduction pathways.

#### Toll pathway

The Toll pathway is a central signaling cascade that regulates the expression of immune-related genes following pathogen recognition by TLRs.

#### Imd pathway

The Imd pathway mediates responses to Gram-negative bacteria, leading to the production of AMPs.

#### JAK/STAT pathway

This pathway is involved in antiviral responses and regulates the expression of genes associated with immune defense.

### Advances in molecular techniques

Recent advancements in molecular biology have significantly enhanced our understanding of crab immunity.

### RNA sequencing and proteomics

These techniques have facilitated the identification and characterization of numerous immune-related genes and proteins, providing a comprehensive understanding of the immune landscape in freshwater crabs.

### Gene editing and rna interference

Emerging tools like CRISPR-Cas9 and RNA interference (RNAi) enable functional studies of immune genes, allowing researchers to dissect the roles of specific components in the immune response (Chomczynski and Sacchi, 1987; Livak and Schmittgen, 2001).

### Implications for aquaculture

Understanding the immune mechanisms of freshwater crabs has practical implications for aquaculture. Enhancing disease resistance through selective breeding or immunostimulants can improve the sustainability and productivity of crab farming (Shields, 2003).

### Future Prospective

Future research should focus on:

1. Investigating the effects of environmental stressors on immune competence.
2. Exploring the potential of immunomodulators in enhancing disease resistance.

3. Conducting comparative studies across different crab species to understand the evolution of immune strategies.

### CONCLUSION

This review highlights the complexity and sophistication of the immune defense mechanisms in the hemolymph of freshwater crabs. Continued research in this field will not only advance our understanding of invertebrate immunity but also support the development of more resilient aquaculture practices as well as This comprehensive review provides an overview of the molecular immune mechanisms in freshwater crabs, with a focus on hemocyte functions, pathogen recognition, and the role of humoral responses. It also highlights recent advancements in molecular techniques that have enabled these discoveries and discusses their implications for aquaculture.

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### CONFLICT OF INTERESTS

The authors declare no conflict of interest

### ETHICS APPROVAL

Not applicable

### REFERENCES

- Amparyup, P., Kondo, H., Hirono, I., Aoki, T., & Tassanakajon, A. (2008). Molecular cloning, genomic organization and recombinant expression of a crustin-like antimicrobial peptide from black tiger shrimp *Penaeus monodon*. *Molecular Immunology*, 45(4), 1085–1093. <https://doi.org/10.1016/j.molimm.2007.07.031>
- Anbuchezen R.M., Ravichandran, S. (2009). Influence of crab haemolymph on clinical pathogens. *Advanced Biology*; 3(4), 104–109.
- Ariki, S., Koori, K., Osaki, T., Motoyama, K., Inamori, K., & Kawabata, S. (2004). A serine protease zymogen functions as a pattern-recognition receptor for lipopolysaccharides. *Proceedings of the National Academy of Sciences of the United States of America*, 101(4), 953–958. <https://doi.org/10.1073/pnas.0306904101>
- Cerenius, L., & Söderhäll, K. (2004). The prophenoloxidase-activating system in invertebrates. *Immunological Reviews*, 198, 116–126. <https://doi.org/10.1111/j.0105-2896.2004.00116.x>

- Cerenius, L., Lee, B. L., & Söderhäll, K. (2008). The proPO-system: pros and cons for its role in invertebrate immunity. *Trends in Immunology*, 29(6), 263–271. <https://doi.org/10.1016/j.it.2008.02.009>
- Cerenius, L., Liang, Z., Duvic, B., Keyser, P., Hellman, U., Palva, E. T., Iwanaga, S., & Söderhäll, K. (1994). Structure and biological activity of a 1,3-beta-D-glucan-binding protein in crustacean blood. *The Journal of Biological Chemistry*, 269(47), 29462–29467.
- Chen, S. C., Yen, C. H., Yeh, M. S., Huang, C. J., & Liu, T. Y. (2001). Biochemical properties and cDNA cloning of two new lectins from the plasma of *Tachypleus tridentatus*: *Tachypleus* plasma lectin 1 and 2+. *The Journal of Biological Chemistry*, 276(13), 9631–9639. <https://doi.org/10.1074/jbc.M008414200>
- Chomczynski, P., & Sacchi, N. (1987). Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Analytical biochemistry*, 162(1), 156–159. <https://doi.org/10.1006/abio.1987.9999>
- Hoffmann, J. A., Kafatos, F. C., Janeway, C. A., & Ezekowitz, R. A. (1999). Phylogenetic perspectives in innate immunity. *Science (New York, N.Y.)*, 284(5418), 1313–1318. <https://doi.org/10.1126/science.284.5418.1313>
- Inamori, K., Ariki, S., & Kawabata, S. (2004). A Toll-like receptor in horseshoe crabs. *Immunological Reviews*, 198, 106–115. <https://doi.org/10.1111/j.0105-2896.2004.0131.x>
- Iwanaga, S., & Lee, B. L. (2005). Recent advances in the innate immunity of invertebrate animals. *Journal of Biochemistry and Molecular Biology*, 38(2), 128–150. <https://doi.org/10.5483/bmbrep.2005.38.2.128>
- Johansson, M. W., Keyser, P., Sritunyalucksana, K., Söderhäll, K. (2000). Crustacean haemocytes and haematopoiesis. *Aquaculture*, 191, (1–3), 45–52.
- Kawabata, S.I., Mutua, T., Iwanaga, S. (1996). The clotting cascade and defense molecules found in the hemolymph of the horseshoe crab. New direction. In: Soderhall K, Iwanaga S, Vasta GR., (eds) *Invertebrate Immunology*. Fair Haven, CT(SOS), 255–283.
- Khoo, L., Robinette, D. W., Noga, E. J. (1999). “Callinectin, an antibacterial peptide from blue crab, *Callinectes sapidus*, hemocytes,” *Marine Biotechnology*, 1(1), 44–51.
- Kobayashi, Y., Takahashi, T., Shibata, T., Ikeda, S., Koshihara, T., Mizumura, H., Oda, T., & Kawabata, S. (2015). Factor B Is the Second Lipopolysaccharide-binding Protease Zymogen in the Horseshoe Crab Coagulation Cascade. *The Journal of Biological Chemistry*, 290(31), 19379–19386. <https://doi.org/10.1074/jbc.M115.653196>
- Li, C., Shields, J. D., & Small, H. J. (2006). Histopathology of parasitic dinoflagellate infections in the blue crab, *Callinectes sapidus*, from Chesapeake Bay. *Diseases of Aquatic Organisms*, 70(3), 235–243.
- Livak, K. J., & Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods (San Diego, Calif.)*, 25(4), 402–408. <https://doi.org/10.1006/meth.2001.1262>
- Muta, T., Seki, N., Takaki, Y., Hashimoto, R., Oda, T., Iwanaga, A., Tokunaga, F., & Iwanaga, S. (1995). Purified horseshoe crab factor G. Reconstitution and characterization of the (1->3)-beta-D-glucan-sensitive serine protease cascade. *The Journal of Biological Chemistry*, 270(2), 892–897.
- Noga, E.J., Arroll, T.A., Fan, Z. (1996). Specificity and some physicochemical characteristics of the antibacterial activity from blue crab *Callinectes sapidus*. *Fish and Shellfish Immunology*, 6(6), 403–412.
- Ratcliffe, N. A., Rowley, A. F., Fitzgerald, S. W., Rhodes, C. P. (1985). *Invertebrate Immunity: Basic Concepts and Recent Advances*, Editor(s): G.H. Bourne, *International Review of Cytology*, Academic Press, 97, 183–350.
- Ravichandran, S., Wahidulla, S., D'Souza, L., & Rameshkumar, G. (2010). Antimicrobial lipids from the hemolymph of brachyuran crabs. *Applied Biochemistry and Biotechnology*, 162(4), 1039–1051. <https://doi.org/10.1007/s12010-009-8843-1>
- Rozen, S., & Skaletsky, H. (2000). Primer3 on the WWW for general users and for biologist programmers. *Methods in Molecular Biology (Clifton, N.J.)*, 132, 365–386. <https://doi.org/10.1385/1-59259-192-2:365>
- Shibata, T., Kobayashi, Y., Ikeda, Y., & Kawabata, S. I. (2018). Intermolecular autocatalytic activation of serine protease zymogen factor C through an active transition state responding to lipopolysaccharide. *The Journal of Biological Chemistry*, 293(29), 11589–11599. <https://doi.org/10.1074/jbc.RA118.002311>
- Shields, J. D. (2003). Research on the health and disease of the blue crab, *Callinectes sapidus*. *Bulletin of Marine Science*, 72(2), 491–497.
- Smith, V. J., & Chisholm, J. R. (2001). Antimicrobial proteins in crustaceans. *Advances in Experimental Medicine and Biology*, 484, 95–112. [https://doi.org/10.1007/978-1-4615-1291-2\\_10](https://doi.org/10.1007/978-1-4615-1291-2_10)
- Söderhäll K. (1982). Prophenoloxidase activating system and melanization - a recognition mechanism of arthropods? A review. *Developmental and Comparative Immunology*, 6(4), 601–611.

- Söderhäll, K., & Smith, V. J. (1983). Separation of the haemocyte populations of *Carcinus maenas* and other marine decapods, and prophenoloxidase distribution. *Developmental and Comparative Immunology*, 7(2), 229–239. [https://doi.org/10.1016/0145-305x\(83\)90004-6](https://doi.org/10.1016/0145-305x(83)90004-6)
- Söderhäll, K., Aspán, A., & Duvic, B. (1990). The proPO-system and associated proteins; role in cellular communication in arthropods. *Research in immunology*, 141(9), 896–907. [https://doi.org/10.1016/0923-2494\(90\)90190-a](https://doi.org/10.1016/0923-2494(90)90190-a)
- Sritunyalucksana K, Söderhäll, K. (2000). The proPO and clotting system in crustaceans. *Aquaculture*; 191, 53–69.
- Stewart, J. E., & Zwicker, B. M. (1972). Natural and induced bactericidal activities in the hemolymph of the lobster, *Homarus americanus*: products of hemocyte-plasma interaction. *Canadian Journal of Microbiology*, 18(9), 1499–1509. <https://doi.org/10.1139/m72-229>
- Tonganunt, M., Wongmanee, K., Sartthai, S., Chotigeat, W., and Phongdara A. (2008). “Crustin protein Amk1 from black tiger shrimp (*Penaeus monodon*) inhibits *Vibrio harveyi* and *Staphylococcus aureus*,” *Journal of Science and Technology*, 30(3), 291–296.
- Vargas-Albores, F., Yepiz-Plascencia, G. (2000). Beta glucan binding protein (BGBP) and its role in immune response. *Aquaculture*, 191, 3–21.
- Vasa, G. R., Quesenberry, M., Ahmed, H., & O’Leary, N. (1999). C-type lectins and galectins mediate innate and adaptive immune functions: their roles in the complement activation pathway. *Developmental and Comparative Immunology*, 23(4-5), 401–420. [https://doi.org/10.1016/s0145-305x\(99\)00020-8](https://doi.org/10.1016/s0145-305x(99)00020-8)
- Vázquez, L., Maldonado, G., Agundis, C., Pérez, A., Cooper, E. L., & Zenteno, E. (1997). Participation of a sialic acid-specific lectin from freshwater prawn *Macrobrachium rosenbergii* hemocytes in the recognition of non-self cells. *The Journal of Experimental Zoology*, 279(3), 265–272. [https://doi.org/10.1002/\(sici\)1097-010x\(19971015\)279:3<265::aid-jez7>3.0.co;2-1](https://doi.org/10.1002/(sici)1097-010x(19971015)279:3<265::aid-jez7>3.0.co;2-1)
- Veeruraj, A., Ravichandran, S., Ramesh Kumar, G. (2008). Antibacterial Activity of Crab Haemolymph on Clinical Pathogens. *Trends in Applied Sciences Research*, 3, 174-181.
- Yeaton R. W. (1981). Invertebrate lectins: II. Diversity of specificity, biological synthesis and function in recognition. *Developmental and Comparative Immunology*, 5(4), 535–545. [https://doi.org/10.1016/s0145-305x\(81\)80028-6](https://doi.org/10.1016/s0145-305x(81)80028-6).